

# Package ‘geecc’

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**Type** Package

**Title** Gene Set Enrichment Analysis Extended to Contingency Cubes

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**Description** Use log-linear models to perform hypergeometric and chi-squared tests for gene set enrichments for two (based on contingency tables) or three categories (contingency cubes). Categories can be differentially expressed genes, GO terms, sequence length, GC content, chromosomal position, phylostrata, divergence-strata, ....

**License** GPL (>= 2)

**Depends** R (>= 3.3.0), methods

**Imports** MASS, hypergea (>= 1.3.0), gplots, Rcpp (>= 0.11.3), graphics, stats, utils

**LinkingTo** Rcpp

**SystemRequirements** Rcpp

**Suggests** hgu133plus2.db, GO.db, AnnotationDbi

**biocViews** BiologicalQuestion, GeneSetEnrichment, WorkflowStep, GO, StatisticalMethod, GeneExpression, Transcription, RNASeq, Microarray

**NeedsCompilation** yes

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 geecc-package

*Gene set enrichment for two or three categories*


---

## Description

This package performs gene set enrichment analyses considering two or three categories. Categories might be regulated genes, sequence length, GC content, GO terms, KEGG pathways and so on.

## Author(s)

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## Examples

```
##
## a completely artificial example run
## through the routines of the package
##
R <- 500
#generate R random gene-ids
ID <- sapply(1:R, function(r){paste( sample(LETTERS, 10), collapse="" ) } )
ID <- unique(ID)

#assign artificial differentially expressed genes randomly
category1 <- list( deg.smallFC=sample(ID, 100, rep=FALSE),
deg.hughFC=sample(ID, 100, rep=FALSE) )
#assign artificial GO terms of genes randomly
category2 <- list( go1=sample(ID, 50, replace=FALSE),
go2=sample(ID, 166, replace=FALSE),
go3=sample(ID, 74, replace=FALSE),
go4=sample(ID, 68, replace=FALSE) )
#assign artificial sequence length of genes randomly
LEN <- setNames(sample(seq(100, 1000, 100), length(ID), replace=TRUE), ID)
category3 <- split( ID, f=factor(LEN, levels=seq(100, 1000, 100)) )
CatList <- list(deg=category1, go=category2, len=category3)

ConCubFilter.obj <- new("concupfilter", names=names(CatList))
ConCub.obj <- new("concup", categories=CatList)
ConCub.obj.2 <- runConCub( obj=ConCub.obj, filter=ConCubFilter.obj, nthreads=1 )
ConCub.obj.3 <- filterConCub( obj=ConCub.obj.2, filter=ConCubFilter.obj )
plotConCub( obj=ConCub.obj.3, filter=ConCubFilter.obj )
x <- getTable(ConCub.obj.3)
```

---

 concub-class

*Class "concup"*


---

## Description

An object of type concub

## Details

Specifying the background population is crucial for the tests for association between factors. Usually the population is the set of all probe sets represented on a micro-array or the set of all genes in a genome. If an expression set is passed with the population-parameter, all probe sets beginning with the pattern "AFFX" (Affymetrix quality control) are removed.

## Objects from the Class

Objects can be created by calls of the form `new("concup", ...)`.

## Slots

`categories`: A named list of named lists. Each item of the outer list represents the two or three categories. Each item of the inner lists represents a variable of the category.

`population`: A character vector containing the background population. As an alternative, an object with class 'eSet', 'ExpressionSet', or 'DGEList'; background population is then set to the outcome of `rownames(population)`.

`keep.empty.vars`: A boolean list with names being names of categories.

`options`: Additional options for individual categories. See Details.

`approx`: specifies the minimum expected value when an exact hypergeometric test (below) or the chi-squared approximation should be used. Defaults to 0.

`null.model`: A formula specifying the null-model of the test.

`test.result`: A list to store test results. Filled up after `runConCub`.

`test.result.filter`: A list to store filtered test results. Filled up after `filterConCub`.

`test.result.filter.heatmap`: A list to store heatmaps for further manipulation. Filled up after `plotConCub`.

The last three slots are not set by the user.

## Methods

**getTable** `signature(object = "concup")`: creates a table containing the results of the tests.

Usage: `getTable(object, na.rm=TRUE, dontshow=list())`

Arguments:

**object** an object of type `concup`

**na.rm** logical. If TRUE (the default), rows with NA P-values (or odds ratios) are removed

The resulting table is a data frame with 8 or 10 columns, depending on if a two- or three-way test was applied

1. 'factor1': this column has the name of the first category
2. 'factor2': this column has the name of the second category
3. 'factor3': this column has the name of the third category
4. n.'factor1': number of items in variable of first category
5. n.'factor2': number of items in variable of second category
6. n.'factor3': number of items in variable of third category
7. p.value: (probably adjusted) p-value
8. log2.odds.ratio: log2 of the sample odds ratio
9. n.tags: number of items at position  $x_{1,1,1}$  or  $x_{1,1}$
10. tags: items at position  $x_{1,1,1}$  or  $x_{1,1}$  (e.g. gene identifiers)

You have to run `filterConCub()` before you can get the table. If `filterConCub()` was not run, a warning is shown and `getTable` returns `NULL`.

```
show signature(object = "concu")
```

### See Also

`GOStats` to perform a simple two-way enrichment analysis

### Examples

```
showClass("concu")
```

---

```
concufilter-class      Class "concufilter"
```

---

### Description

An object of type `concufilter`

### Details

The large number of different filter options (and corresponding getter and setter accessors) makes it necessary to maintain them in a special class. This differs from other packages like `GOstats`, where arguments for controlling the program and the results are stored in the same object.

### Objects from the Class

Objects can be created by calls of the form `new("concufilter", ...)`.

### Slots

For a more detailed description of some slots see below.

`nfact`: a numeric giving the number of factors

`names`: a character vector giving the name of each factor

`p.value`: a numeric giving the P-value to be considered. Defaults to 0.1.

`test.direction`: a character giving the direction of association. Defaults to "two.sided".

`minimum.l2or`: a numeric giving the minimum absolute log<sub>2</sub> odds ratio to be considered. Defaults to 0.

`skip.min.group`: a numeric giving the minimum number of tag a group is allowed to have. Defaults to 2.

`skip.min.obs`: a numeric giving the minimum number at the position of interest allowed. Defaults to 2.

`skip.zeroobs`: a logical. Defaults to `TRUE`.

`drop.insignif.layer`: A vector of logicals. By default, all positions are set to `FALSE`.

`drop.wrongdir.layer`: A vector of logicals. By default, all positions are set to `FALSE`.

`drop.lowl2or.layer`: A vector of logicals. By default, all positions are set to `FALSE`.

## Methods and slot accessors

Several methods are implemented for class `concufilter`. They can be roughly grouped into informative, basic, skip-test, and data-reduction methods.

Individual options can be accessed by the corresponding getter and setter methods, for instance

**skip.zerooks** signature(`x = "concufilter"`): get current setting to skip test in case of zero cell

**skip.zerooks<-** signature(`x = "concufilter"`): set a new value to skip test in case of zero cell

**Informative methods:** Currently only a single method is implemented.

**show** signature(`object = "concufilter"`): a short summary about current filter settings

### Basic filters/thresholds:

**p.value** The maximum P-value that should be taken into account

**minimum.l2or** The minimum absolute of log2 odds ratio that should be taken into account

**test.direction** The direction of the association. Can be "two.sided", "greater" (test for over-representation), or "less" (test for under-representation)

**Skip test:** The following filters cause a skip of a test, i.e. the test is never run if at least one of the conditions is fulfilled.

**skip.zerooks** skip the test, if the position of interest ( $x_{00}$  or  $x_{000}$ ) is zero, i.e. no tag from the population matches the conditions defined at the marginals.

**skip.mingroup** skip the test, if the groups considered at the position of interest are too small at all.

**skip.minobs** skip the test, if the position of interest contains less than 'this value' entries.

**Data reduction:** The following filters reduce the amount of outcome of the tests. They are applied to both, the (2 or 3 dimensional) table containing the odds ratios and the table containing the corresponding P-values.

**drop.insignif.layer** drop all layers in the tables where all  $P$ -values are greater than the value defined in 'p.value'.

**drop.wrongdir.layer** drop all layers in the tables where all odds ratios are showing into the opposite direction as defined in 'test.direction'.

**drop.lowl2or.layer** drop all layers in the tables where all absolutes of the log2 odds ratios are smaller than 'min.l2or'.

## Examples

```
showClass("concu")
```

---

filterConCub	<i>Filter results from two- or three-way tests</i>
--------------	--

---

### Description

Performs filtering on results from two- or three-way tests

### Usage

```
filterConCub(obj, filter, p.adjust.method = "none", verbose=1)
```

### Arguments

obj	object of type concub
filter	object of type concubfilter
p.adjust.method	set adjustment-method for p-values. Must match any of <a href="#">p.adjust.methods</a> .
verbose	An integer specifying the level of verbosity.

### Details

You have to execute [runConCub](#) before filtering.

### Value

an (extended) object of type concub with filtered test results

### Examples

```
# a character vector listing possible
# adjustment approaches
p.adjust.methods
```

---

GO2list	<i>Filter GO and KEGG database</i>
---------	------------------------------------

---

### Description

Filter GO and KEGG database and transform database to list

### Usage

```
GO2list(dbase, go.cat = NULL, rm = NULL, keep = NULL)
KEGG2list(dbase, rm = NULL, keep = NULL)
GO2offspring(x)
GO2level(x, go.level=-1, relation=c("is_a"))
```

## Arguments

dbase	A datastructure storing identifiers of GO/KEGG terms and assigned genes. Can be one of  <b>database</b> usually of class ‘ProbeGo3AnnDbBimap’ (as defined in package “AnnotationDbi”) <b>named list</b> with keys being the identifiers and values being genes <b>dataframe</b> with first column being the identifiers and second column being genes. Additional columns are ignored.
x	a list with keys being the identifiers and values being genes (e.g. output of GO2list)
go.cat	GO category ("MF", "BP", "CC") that should be returned and filtered
go.level	Level in the DAG of GO terms. Defaults to “-1” for pass through without modification. Otherwise: a positive integer giving the level at which GO terms should be grouped together.
rm	remove these terms
keep	keep only these terms
relation	relationships in GO hierarchy that should be considered. Defaults to “is_a”

## Details

The settings for “rm” and “keep” can be combined, allowing for efficient reduction of the number of GO terms and KEGG pathways, respectively.

Providing a named list instead of a database can be useful for non-model organisms, where only a draft Blast2GO-annotation is available. In this case, the names of the list are the GO terms (or KEGG pathways) and the content of each list item is a character vector with tag-ids.

The function GO2offspring does the same as the databaseGO2ALLPROBES function does (e.g. hgu133plus2GO2ALLPROBES). I.e. instead of representing only features (probe sets, genes, ...) assigned to the GO terms directly, it also contains all features assigned to all children (offsprings).

The function GO2level groups GO terms together at a more general level to simplify data interpretation and speed up runtime. This function works according to the level option provided by DAVID, but the number of levels is not restricted.

## Value

A named list with each slot containing the ids for the term or pathway.

## Examples

```
library(hgu133plus2.db)
x <- GO2list(dbase=hgu133plus2GO2PROBE, go.cat="CC",
rm=c("GO:0000139", "GO:0000790", "GO:0005730", "GO:0005739"))
```

---

marioni

*Affymetrix microarray gene expression data*

---

### Description

The experiment aims to detect differentially expressed genes in Affymetrix micro-arrays and RNA-seq data in a comparative study. For this, samples from two tissues (liver and kidney) were compared.

### Usage

```
marioni
```

### Format

A `data.frame` containing gene expression values from an Affymetrix microarray, including P-values,  $\log_2$ -fold changes and alternative annotations

### Value

A `data.frame`.

### Source

<http://giladlab.uchicago.edu/data.html>

### References

Marioni, J. C. et al. (2008) RNA-seq: an assessment of technical reproducibility and comparison with gene expression arrays. *Genome research*

### Examples

```
data(marioni)
head(marioni[, 1:5])
```

---

plotConCub

*Generate a heatmap showing  $\log_2$  odds ratios and P-values.*

---

### Description

The function generates a heatmap by calling the `heatmap.2`-function from the `gplots`-package. Each cell shows the  $\log_2$  odds ratio of the test for the corresponding variables. In addition, stars indicate the *P*-value for this test.

### Usage

```
plotConCub(obj, filter, fix.cat = 1, show=list(), dontshow=list(),
  args_heatmap.2 = list(), col = list(range = NULL),
  alt.names = list(), t = FALSE)
```



**Arguments**

obj	An object with class concub
filter	An object with class concubfilter
fix.cat	The heatmap can only visualize a two-dimensional table. In case of three-dimensions, one dimension (category) must be fixed.
show	A named list. The names are the names of the categories. Each item is a character vector of variables that should be shown in the plot.
dontshow	A named list. The names are the names of the categories. Each item is a character vector of variables that should not be shown in the plot.
args_heatmap.2	Arguments passed to <code>'heatmap.2'</code> . Can be used to change size of fonts etc.
col	A vector of colors, for instance from <a href="#">heat.colors</a>
alt.names	Substitute variables by alternative terms. For instance, if variables are artificial ids, they can be substituted by descriptive text for the heatmap.
t	logical; transpose matrix for heatmap. Default FALSE.

**Value**

An object with class concub.

**Examples**

```
##
## a completely artificial example run
## through the routines of the package
##
R <- 500
#generate R random gene-ids
ID <- sapply(1:R, function(r){paste( sample(LETTERS, 10), collapse="" ) } )
ID <- unique(ID)

#assign artificial differentially expressed genes randomly
category1 <- list( deg.smallFC=sample(ID, 100, rep=FALSE),
deg.hughFC=sample(ID, 100, rep=FALSE) )
#assign artificial GO terms of genes randomly
category2 <- list( go1=sample(ID, 50, replace=FALSE),
go2=sample(ID, 166, replace=FALSE),
go3=sample(ID, 74, replace=FALSE),
go4=sample(ID, 68, replace=FALSE) )
#assign artificial sequence length of genes randomly
LEN <- setNames(sample(seq(100, 1000, 100), length(ID), replace=TRUE), ID)
category3 <- split( ID, f=factor(LEN, levels=seq(100, 1000, 100)) )
CatList <- list(deg=category1, go=category2, len=category3)

ConCubFilter.obj <- new("concubfilter", names=names(CatList))
ConCub.obj <- new("concub", categories=CatList)
ConCub.obj.2 <- runConCub( obj=ConCub.obj, filter=ConCubFilter.obj, nthreads=1 )
ConCub.obj.3 <- filterConCub( obj=ConCub.obj.2, filter=ConCubFilter.obj )
plotConCub( obj=ConCub.obj.3, filter=ConCubFilter.obj )
```

---

pval2star                      *Transform P-values to stars*

---

**Description**

Transform P-values to stars

**Usage**

```
pval2star(x)
```

**Arguments**

x                      A matrix of P-values

**Details**

Use stars as simplification of P-values

**Value**

A character matrix of same dimension and names as x with stars instead of P-values.

**Examples**

```
x <- matrix( runif(25), nrow=5, dimnames=list(LETTERS[1:5], letters[1:5]) )
pval2star(x)
```

---

runConCub                      *Enrichment analysis on two- or three-way contingency tables.*

---

**Description**

Perform the enrichment analysis on two- or three-way contingency tables.

**Usage**

```
runConCub(obj, filter, nthreads = 2, subset = NULL,
  verbose=list(output.step=0, show.cat1=FALSE,
  show.cat2=FALSE, show.cat3=FALSE))
```

**Arguments**

obj                      an object with class concub  
 filter                      an object with class concubfilter  
 nthreads                      number of threads to use in hypergeom. test  
 subset                      a named list. Restrict enrichment analysis to these category variables  
 verbose                      A list to control verbosity:

**output.step:** after how many variables passed of category 2 a control output should be printed

**show.cat1:** show current level of category 1

**show.cat2:** show current level of category 2

**show.cat3:** show current level of category 3

## Details

This function applies a test for association for all combinations of all variables of all categories to be tested. Depending on the settings in the concubfilter-object, a one-sided or two-sided test is made, using the exact hypergeometric test as implemented in the hypergea-package if the smallest expected value is smaller than 5, or using the chi-squared test as implemented in the loglm-function implemented in the MASS-package. The minimum expected value can be changed in the concub-object by the user (approx-parameter). In this function only filter-settings those filter settings are used, which skip the tests.

## Value

An object with class concub.

## Examples

```
##
## a completely artificial example run
## through the routines of the package
##
R <- 500
#generate R random gene-ids
ID <- sapply(1:R, function(r){paste( sample(LETTERS, 10), collapse="" ) } )
ID <- unique(ID)

#assign artificial differentially expressed genes randomly
category1 <- list( deg.smallFC=sample(ID, 100, rep=FALSE),
deg.hughFC=sample(ID, 100, rep=FALSE) )
#assign artificial GO terms of genes randomly
category2 <- list( go1=sample(ID, 50, replace=FALSE),
go2=sample(ID, 166, replace=FALSE),
go3=sample(ID, 74, replace=FALSE),
go4=sample(ID, 68, replace=FALSE) )
#assign artificial sequence length of genes randomly
LEN <- setNames(sample(seq(100, 1000, 100), length(ID), replace=TRUE), ID)
category3 <- split( ID, f=factor(LEN, levels=seq(100, 1000, 100)) )
CatList <- list(deg=category1, go=category2, len=category3)

ConCubFilter.obj <- new("concubfilter", names=names(CatList))
ConCub.obj <- new("concub", categories=CatList)
ConCub.obj.2 <- runConCub( obj=ConCub.obj, filter=ConCubFilter.obj, nthreads=1 )
ConCub.obj.2
```

---

`sortAscii`*Optimized operations of sets of character-vectors*

---

**Description**

Sort and use pre-sorted character vectors in set-operations

**Usage**

```
sortAscii(x)
intersectPresort(pop, x)
setdiffPresort(pop, x)
```

**Arguments**

<code>x</code>	an unsorted vectors of strings
<code>pop</code>	a sorted vector of strings

**Details**

By default, sorting is done lexicographically in R. The routine `sortAscii` does sorting according to the ASCII-order as done in C/C++.

For routines `intersectPresort` and `setdiffPresort` the first argument has to be sorted according to ASCII-order. This first argument is expected to be large compared to the second argument. Both functions are wrappers for optimized C++-functions performing the set-operation.

**Value**

An character-vector. In case of `intersectPresort` and `setdiffPresort`, these vectors are unnamed.

**Examples**

```
AA <- matrix( sample( c(LETTERS, letters), 10*30000, rep=TRUE ), ncol=10 )
A <- unique(apply(AA, 1, paste, collapse=""))
B <- sample(AA, 100, replace=FALSE); B <- c(B, "1234")

res <- intersectPresort( sortAscii(A), B )
```

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